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### First report of the *Brevipalpus*-transmitted (Trombidiformes: Tenuipalpidae) virus, *Orchid fleck virus* (Mononegavirales: Rhabdoviridae) infecting three ornamentals in Florida

Austin **Fife**1, Daniel **Carrillo**2, Gary **Knox**3, Fanny **Iriarte**4, Kishore **Dey**5, Avijit **Roy**6, Ronald **Ochoa**7, Gary **Bauchan**8 , Mathews **Paret**4, Xavier **Martini**1\*

1 University of Florida, Department of Entomology and Nematology, North Florida Research and Education Center, Quincy FL 32351

2 University of Florida, Department of Entomology and Nematology, Tropical Research and Education Center, Homestead FL 33031

3 University of Florida, Department of Environmental Horticulture, North Florida Research and Education Center, Quincy FL 32351

4 University of Florida, Department of Plant Pathology, North Florida Research and Education Center, Quincy FL 32351

5 The Florida Department of Agriculture and Consumer Services, Division of Plant Industry, Section of Plant Pathology, Doyle Conner Building, 1911 SW 34th street, Gainesville, FL 32608

6 United States Department of Agriculture – Agriculture Research Service, Molecular Plant Pathology Laboratory, 10300 Baltimore Ave, Bldg. 4 BARC-West, Beltsville, MD 20705

7 United States Department of Agriculture - Agriculture Research Service, Systematic Entomology Laboratory 10300 Baltimore Ave, Bldg. 5 BARC-West, Beltsville, MD 20705

8 United States Department of Agriculture - Agriculture Research Service, Electron and Confocal Microscopy Unit, Bldg. 12 BARC-West, 10300 Baltimore Ave, Beltsville, MD 20705

\*Corresponding author; E-mail: xmartini@ufl.edu Phone: 850-875-7160 Fax: 352-846-6617

### Abstract

Several flat mite species, all from the genus *Brevipalpus* Donnadieu (Trombidiformes: Tenuipalpidae), are the only known vectors of dichorhaviruses. The *B. californicus* species group exclusively can transmit *Orchid fleck dichorhavirus* (OFV) in a persistent propagative manner. OFV is the type species for the genus *Dichorhavirus* and infects more than fifty plant species belongs to the family Orchidaceae, Asparagaceae (Nolinoidaea), and Rutaceae (*Citrus*). During June 2020, chlorotic ringspot symptoms on Giant Lilyturf (*Liriope* spp., cv. ‘Gigantea’) were observed in a landscape in Leon County, Florida. Similar symptoms were observed on XXX in Alachua County, Florida. In both cases, the presence of OFV was confirmed using OFV specific conventional reverse transcription polymerase chain assay (RT-PCR) assay and Sanger sequencing. RT-PCR amplicons had a 98% identity with the known OFV sequences available in the Genbank. The identification was also confirmed with the quantitative RT-PCR (RT-qPCR). Additional samples were taken from other Nolinoidaea, including *Liriope* *muscari*, *Ophiopogon japonicus*, *O. intermedius* and *Aspidistra elatior* Blume (Asparagaceae: Nolinoidaea) in Leon and Alachua counties. Identification of partial genome sequence confirmed the presence of both the orchid strains (OFV-Orc1 and OFV-Orc2) in Florida. Three mite species were recovered from OFV-infected plants: *Brevipalpus californicus* sensu lato, *B. obovatus* Donnadieu and *B. confusus* Banks. One of these species is presumably responsible for OFV transmission. Florida has various mite species of *Brevipalpus* and a diverse array of susceptible native and introduced plant species in the landscape. We suggest that OFV already has a wide distribution in Florida which will continue to spread if unchecked, representing a potential threat for *Liriope* spp., *Ophiopogon spp.* and *Aspidistra elatior* which are commonly used in landscaping in Southeastern US. In this study we are reporting three new hosts from the family Asperagaceae. To know the extended host range of OFVs, a survey in Florida's citrus growing regions is essential, emphasizing plants within the families Rutaceae and Asparagaceae.

### Keywords:

False spider mite, flat mite, *Brevipalpus*-transmitted viruses, *Liriope*, Nolinoideae, *Ophiopogon*, Ruscaceae, Rutaceae, Asparagaceae, orchid, Orchidaceae, pests, ornamental plants, orchid fleck virus.

*Orchid fleck dichorhavirus* (OFV) is the type member for the genus *Dichorhavirus*, family Rhabdoviridae; a bacilliform, nuclear rhabdoviruses composed of two segments of single-stranded, negative-sense RNA which infects plants (Dietzgen et al. 2014, Walker et al. 2018, Amarasinghe et al. 2019). Other members of this genus are: *Citrus chlorotic spot virus*, *Citrus leprosis virus* N, *Clerodendrum chlorotic spot virus* and *Coffee ringspot virus* (Dietzgen, Freitas-Astúa, et al. 2018). Flat mites from the genus *Brevipalpus* Donnadieu (Trombidiformes: Tenuipalpidae) are the only known vectors for dichorhaviruses (Maeda 1998), and mites in the *Brevipalpus californicus* (Banks) group the only known to transmit OFV in a persistent propagative manner (Kondo et al. 2003).

OFV-infected plants exhibit various symptoms depending on the infected plant species as well as the strain of the OFV (Kubo et al. 2009), but symptoms typically appear as chlorotic flecks, which ultimately coalesce into larger spots or ringspot patterns (Fig. 1).

OFV was first described infecting *Cymbidium* orchids in Japan (Doi et al. 1977). OFV and OFV-like rhabdoviruses have been reported infecting orchids in Asia: [China (Peng et al. 2017), Korea Zheng et al. (2013)], Africa: [South Africa (Blanchfield et al. 2001)], North America: [The United States (Blanchfield et al. 2001, Bratsch et al. 2015), Mexico (Otero et al. 2021)], South America: [Brazil (Kitajima et al. 1974, Kitajima et al. 2001), Colombia (Kubo et al. 2009), Costa Rica (Freitas-Astúa et al. 2002), Paraguay (Ramos-González et al. 2015)], Europe: [Denmark (Begtrup 1972), France (Sauvêtre et al. 2018), Germany (Petzold 1971, Lesemann and Doraiswamy 1975)] and Oceania: [Australia (Lesemann and Begtrup 1971, Lesemann and Doraiswamy 1975, Gibbs 2000), Fiji (Pearson et al. 1993), Vanuatu (Pearson et al. 1993)]. The prevalence of OFV and its mite vector is thought to be associated with the importation of infected orchids (Dietzgen, Freitas-Astúa, et al. 2018).

OFV naturally infects more than fifty species of Orchidaceae (Kitajima et al. 2010, Kondo et al; 2006, Peng et al. 2013), some Asparagaceae (Nolinoidaea) Dietzgen, Tassi, et al. (2018), as well as Rutaceae (*Citrus*), where it causes citrus leprosis-like symptoms (Cook et al; 2019, Olmedo-Velarde et al; 2021 and Roy et al. 2015, 2020). Mechanical transmission of OFV is possible under lab conditions to various Chenopodiaceae, Aizoaceae, Fabaceae, and Solanaceae (Chang et al. 1976, Kondo et al. 2003, Peng et al. 2013).

During June 2020, chlorotic ringspot symptoms were observed on Giant Lilyturf *Liriope* spp., cv. ‘Gigantea’ in a landscape of Leon County, Florida (Fig. 1). *Liriope* belong to a group of plants in the family Asparagaceae, subfamily Nolinoidaea, which includes a diverse array of various monocotyledonous liliod plants which are native to southeastern Asia (Chase et al. 2009, Meng et al. 2021). *Liriope* and the closely related *Ophiopogon* Ker Gawler (Asparagaceae: Nolinoidaea) are considered the most important ground cover sold by the nursery industry in southeastern US (Mcharo et al. 2003).

Viral infections of suspected plant leaf samples were initially tested at the Plant Disease Diagnostic Clinic at the North Florida Research and Education Center (NFREC) in Quincy, FL. All the samples tested negative for begomovirus, potyvirus, tospovirus as well as for Impatiens necrotic spot virus, Tobacco mosaic virus and Tomato spotted wilt virus. The infected materials were subsequently sent to the Florida Department of Agriculture and Consumer Services (FDACS). The presence of OFV was confirmed using OFV generic R2-Dicho-GF and R2-Dicho-GR primers (Roy et al. 2020) by one step conventional RT-PCR, amplifying ~800 nt of L-gene (RNA2) amplicon from an infected *Liriope* leaf sample. The RT-PCR amplicon was sequenced using Sanger sequencing and confirmed the presence of OFV in the tested samples. Sanger sequencing of RT-PCR amplicons shared 98% nucleotide identity with orchid strains of OFV (GenBank Accession numbers: AB244418 and LC222630) (Kondo et al. 2006, 2017). Further surveys of putatively OFV-infected plants were taken during subsequent visits to the initial site of collection. Plant samples included various cultivars of Nolinoid plants, including *Liriope* spp., *Ophiopogon* spp., and *Aspidistra elatior* Blume (Asparagaceae: Nolinoidaea) which were located at the same site as the original infected *Liriope* spp., cv ‘Gigantea.’ *A. elatior* infected with OFV have chlorotic leaves or chlorotic flecks. No ringspots have been observed on *A. elatior* to date. All plant samples were tested via RT-PCR at the NFREC to confirm the presence of OFV.

Further surveys of plants belonging to the subfamily Nolinoidaea in Florida have revealed more sites with symptomatic plants in Leon and Alachua counties. Mites were collected from symptomatic plants in Leon county and observed with phase contrast microscopy. We encountered both an unidentified eriophyoid mite species on the *Liriope* spp., along with flat mites on all the plant species tested (*Liriope* spp.*, Ophiopogon* spp., and *A. elatior*). The flat mites were originally identified as *Brevipalpus californicus* (Banks) sensu lato by the first author and later confirmed by FDACS via Differential Interference Contrast (DIC) microscopy. *Brevipalpus* mites have been previously associated with OFV (Dietzgen et al. 2018, García-Escamilla et al. 2018, Beltran-Beltran et al. 2020) as well as similar diseases (Kitajima et al. 2010) and are known to feed on a large variety of economically important plants (Childers et al. 2003, Akyazi et al. 2017). The *Brevipalpus* mite species complex is known to contain cryptic species (Childers and Rodrigues 2011) that require advanced microscopy techniques, such as cryo scanning electron microscopy (Cryo-SEM) for species identification (León and Nadler 2010, Skoracka et al. 2015, Beard et al. 2015). Additional mite samples from the original OFV detection site were observed with Cryo-SEM. The determinations agreed with both prior identifications of *B. californicus* s.l. but revealed the presence of two other species *B. obovatus* and *B. confusus.*

The first report of OFV in the United States is thought to be Ko et al. (1985), who describes nuclear inclusions caused by an undescribed bacilliform rhabdovirus in *Brassia* orchids. The significance of this report is their reference to spoke-wheel configurations of the viral particles (Ko et al. 1985), a sign typically associated with OFV infection (Chang et al. 1976). Unfortunately, Ko et al. (1985) made no mention of mites or further investigations of this virus. The first certain report of OFV was in Hawaii in 2001 and in the continental US in 2015 (Bratsch et al. 2015). The authors confirmed the presence of OFV in *Phalaenopsis* hybrids in the United States, using TEM of ultrathin sections of plant tissue as well as molecular sequence analysis and its association with *Brevipalpus* mites. The authors did not make a conclusive species identification but suggested the mite vector was within the *B. californicus* group, as referred by Kondo et al. (2003).

OFV has been reported in other Nolinoidaea in Australia (Mei et al. 2016, Dietzgen et al. 2018), including *Liriope spicata* (Thunb.) Lour. (Mei et al. 2016). The Florida collected plants of *Liriope* spp., cv 'Gigantea' are thought to belong to either *Liriope muscari* or *Liriope gigantea.* We are not aware of any previous report of OFV infection in *Ophiopogon* plants. Unfortunately, the Ophiopogonae group includes species of *Liriope* and *Ophiopogon* which are very similar in appearance and growth habit, with few useful morphological characters used for their classification (Fantz 2008a). Furthermore, the horticultural industry has created a diverse array of cultivars of these plants, which are often mislabeled (Fantz 2008a). Aside from the taxonomic confusion created by humans, natural hybrids between *Ophiopogon* and *Liriope* have created a natural source of error for reconstructing phylogenies (Zhou et al. 2009). Together, these factors make it difficult to differentiate and identify plants in the landscape by visual inspection alone. These obfuscations of species identity may be accounted for in the future via sequence comparisons of the OFV-infected plants, but these comparisons are beyond the scope of our current report. Nonetheless, we are confident that ours is the first report of OFV infecting *A. elatior*; although (Zheng et al. 2013) mentions the association of *B. californicus* with *A. elatior* they make no mention of OFV symptoms in this plant.

### Conclusion

OFV is the type member of the genus *Dichorhavirus* (family *Rhabdoviridae*; order *Mononegavirales*) contain a bipartite, single-stranded and negative sense RNA genome. The dichorhavirus that infects citrus in Hawaii, Mexico, Colombia, and South Africa are identical to the OFV in gene order, content, and the genome sequence. According to the International committee on Taxonomy of Viruses (ICTV) classification, OFV consist of two orchid strains (OFV-Orc1 and OFV-Orc2) and two citrus strains (OFV-Cit1 and OFV-Cit2). The two orchid strains of OFV also infects citrus, but none of the citrus strains have been reported from any orchid species. Detecting OFV in Florida represents a concern for horticulturists who grow orchids, *Liriope*, *Ophiopogon*, or other susceptible Asparagaceae species which are commonly used in landscaping. Florida is also home to a plethora of native and naturalized orchid species, many of which are threatened, including cultivating *Vanilla* in southern Florida and the famous Ghost Orchid, [*Dendrophylax lindenii* (Lindl.) Benth]. ex Rolfe. Citrus leprosis was present in Florida in 1860’s and eradicated in the mid-1960s. In fact, Kitajima et al. (2011) found that the Citrus Leprosis virus (CiLV) which previously affected Florida citrus was a nuclear type of citrus leprosis closely related to OFV strains (Roy et al. 2013). Association of a distant relative of OFV named Citrus leprosis dichorhavirus -N0 (CiLV-N0) was confirmed in relation to the leprosis disease outbreak in Florida (Hartung et al; 2015). The recent detection of OFV-Orc1 in South Africa (Cook et al; 2019) in *C. sinensis* (Navel and Valencia orange) and OFV-Orc2 in Hawaii (Olmedo-Velarde et al; 2021) in *C. reticulata* (mandarin) and *C. jambhiri* (rough lemon) associated with leprosis-like symptoms highlights the threat of different strains of OFV on citrus which will be a definite concern to the multi-billion dollar citrus industry. Similarly, some OFV isolates are known to be involved with citrus leprosis disease in Mexico (Roy et al. 2015), which is a cause for concern for the citrus industry. *B. californicus* and *B. yothersi* are both known vectors of Dichorhaviruses (OFV) and Cileviruses (Citrus Leprosis) (Knorr 1968, Kondo et al. 2003, Beltran-Beltran et al. 2020) and *B. obovatus* is a suspected vector as well (Childers et al. 2003). All three mite species/complexes are present in Florida (Childers et al. 2003, Akyazi et al. 2017). Therefore, it is critical to identify the vector of OFVs in Florida and monitor its spread to determine the risk this virus represents for the native plants, agriculture and the ornamental/landscaping industries of Florida and the surrounding regions.

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### References

**Akyazi, R., E. A. Ueckermann, and O. E. Liburd**. **2017**. New report of *Brevipalpus yothersi* (prostigmata: Tenuipalpidae) on blueberry in Florida. Florida Entomologist. 100: 731–739.

**Alves, E. B., C. Omoto, and C. R. Franco**. **2000**. Resistência cruzada entre o dicofol e outros acaricidas em *Brevipalpus phoenicis* (Geijskes) (acari: tenuipalpidae). Anais da Sociedade Entomológica do Brasil. 29: 765–771.

**Amarasinghe, G. K., M. A. Ayllón, Y. Bào, C. F. Basler, S. Bavari, K. R. Blasdell, T. Briese, P. A. Brown, A. Bukreyev, A. Balkema-Buschmann, U. J. Buchholz, C. Chabi-Jesus, K. Chandran, C. Chiapponi, I. Crozier, R. L. de Swart, R. G. Dietzgen, O. Dolnik, J. F. Drexler, R. Dürrwald, W. G. Dundon, W. P. Duprex, J. M. Dye, A. J. Easton, A. R. Fooks, P. B. H. Formenty, R. A. M. Fouchier, J. Freitas-Astúa, A. Griffiths, R. Hewson, M. Horie, T. H. Hyndman, D. Jiāng, E. W. Kitajima, G. P. Kobinger, H. Kondō, G. Kurath, I. V. Kuzmin, R. A. Lamb, A. Lavazza, B. Lee, D. Lelli, E. M. Leroy, J. Lǐ, P. Maes, S.-Y. L. Marzano, A. Moreno, E. Mühlberger, S. V. Netesov, N. Nowotny, A. Nylund, A. L. Økland, G. Palacios, B. Pályi, J. T. Pawęska, S. L. Payne, A. Prosperi, P. L. Ramos-González, B. K. Rima, P. Rota, D. Rubbenstroth, M. Shı̄, P. Simmonds, S. J. Smither, E. Sozzi, K. Spann, M. D. Stenglein, D. M. Stone, A. Takada, R. B. Tesh, K. Tomonaga, N. Tordo, J. S. Towner, B. van den Hoogen, N. Vasilakis, V. Wahl, P. J. Walker, L.-F. Wang, A. E. Whitfield, J. V. Williams, F. M. Zerbini, T. Zhāng, Y.-Z. Zhang, and J. H. Kuhn**. **2019**. Taxonomy of the order Mononegavirales: Update 2019. Archives of Virology. 164: 1967–1980.

**Argolo, P. S., A. M. Revynthi, M. A. Canon, M. M. Berto, D. J. Andrade, İ. Döker, A. Roda, and D. Carrillo**. **2020**. Potential of predatory mites for biological control of *brevipalpus yothersi* (acari: tenuipalpidae). Biological Control. 149: 104330.

**Baker, E. W., and D. M. Tuttle**. **1987**. The false spider mites of Mexico (tenuipalpidae: acari). (technical report No. 1706). The United States Department of Agriculture - Agricultural Research Service.

**Beard, J. J., R. Ochoa, R. Bauchan G., D. Trice M., J. Redford A., W. Walters T., and C. Mitter**. **2012**. Flat mites of the world edition 2. (<http://idtools.org/id/mites/flatmites/>).

Beard, J.J., Ochoa, R., Braswell, W.E. & Bauchan, G.R., 2015. *Brevipalpus phoenicis* (Geijskes) species complex (Acari: Tenuipalpidae) – a closer look. Zootaxa. 3944,1–67. doi:10.11646/zootaxa.3944.1.1

**Begtrup, J.** **1972**. Structure of a bacilliform virus in Dendrobium as revealed by negative staining. Journal of Phytopathology. 75: 268–273.

**Beltran-Beltran, A. K., M. T. Santillán-Galicia, A. W. Guzmán-Franco, D. Teliz-Ortiz, M. A. Gutiérrez-Espinoza, F. Romero-Rosales, and P. L. Robles-Garcı́a**. **2020**. Incidence of citrus leprosis virus c and orchid fleck dichorhavirus citrus strain in mites of the genus *Brevipalpus* in Mexico. Journal of Economic Entomology. 113: 1576–1581.

**Blanchfield, A. L., A. M. Mackenzie, A. Gibbs, H. Kondo, T. Tamada, and C. R. Wilson**. **2001**. Identification of orchid fleck virus by reverse transcriptase-polymerase chain reaction and analysis of isolate relationships. Journal of Phytopathology. 149: 713–718.

**Bratsch, S. A., B. E. Lockhart, and C. Ishimaru**. **2015**. Confirmation of first report of orchid fleck virus in *phalaenopsis* hybrid orchids in the USA. Plant Health Progress. 16: 146–148.

**Broussard, M. C.** **2007**. A horticultural study of *Liriope* and *Ophiopogon*: Nomenclature, morphology, and culture (PhD thesis).

**Campos, F. J., and C. Omoto**. **2002**. Experimental and Applied Acarology. 26: 243–251.

**Chambers, A. H., P. Moon, V. Edmond, and E. Bassil**. **2019**. Vanilla cultivation in southern Florida. EDIS. 2019: 7.

**Chang, M. U.** **1991**. Studies on the viruses in orchids in Korea. 2. *Dendrobium mosaic virus, Odontoglossum ringspot virus, Orchid fleck virus* and unidentified potyvirus. Korean J Plant Pathol. 7: 118–129.

**Chang, M. U., Arai. Kei, Doi. Yoji, and Yora. Kiyoshi**. **1976**. Morphology and intracellular appearance of *Orchid fleck virus*. Japanese Journal of Phytopathology. 42: 156–157.

**Chase, Mark. W., James. L. Reveal, and M. F. Fay**. **2009**. A subfamilial classification for the expanded asparagalean families amaryllidaceae, asparagaceae and xanthorrhoeaceae. Botanical Journal of the Linnean Society. 161: 132–136.

**Chen, T.-Y., J. V. French, T.-X. Liu, and J. V. da Graça**. **2006**. Predation of *Galendromus helveolus* (acari: Phytoseiidae) on *Brevipalpus californicus* (acari: tenuipalpidae). Biocontrol Science and Technology. 16: 753–759.

**Childers, C. C., and J. C. V. Rodrigues**. **2011**. An overview of *Brevipalpus* (acari: Tenuipalpidae) and the plant viruses they transmit. Zoosymposia. 6: 180–192.

**Childers, C. C., J. C. V. Rodrigues, and W. C. Welbourn**. **2003**. Host plants of *brevipalpus californicus*, *b. Obovatus*, and *b. Phoenicis* (acari: Tenuipalpidae) and their potential involvement in the spread of viral diseases vectored by these mites. Experimental and Applied Acarology. 30: 29–105.

**Cook, G., W. Kirkman, R. Clase, C. Steyn, E. Basson, P. H. Fourie, S. D. Moore, T. G. Grout, E. Carstens, and V. Hattingh**. **2019**. Orchid fleck virus associated with the first case of citrus leprosis-N in South Africa. European Journal of Plant Pathology. 155: 1373–1379.

**Dietzgen, R. G., J. Freitas-Astúa, C. Chabi-Jesus, P. L. Ramos-González, M. M. Goodin, H. Kondo, A. D. Tassi, and E. W. Kitajima**. **2018**. Dichorhaviruses in their host plants and mite vectors, pp. 119–148. *In* Advances in Virus Research. Elsevier.

**Dietzgen, R. G., J. H. Kuhn, A. N. Clawson, J. Freitas-Astúa, M. M. Goodin, E. W. Kitajima, H. Kondo, T. Wetzel, and A. E. Whitfield**. **2014**. Dichorhavirus: A proposed new genus for *Brevipalpus* mite-transmitted, nuclear, bacilliform, bipartite, negative-strand RNA plant viruses. Archives of Virology. 159: 607–619.

**Dietzgen, R. G., A. D. Tassi, J. Freitas-Astúa, and E. W. Kitajima**. **2018**. First report of orchid fleck virus and its mite vector on green cordyline. Australasian Plant Disease Notes. 13.

**Doi, Y., M. U. Chang, and K. Yora**. **1977**. Orchid fleck virus. CMI/AAB descriptions of plant viruses.

**Fantz, P. R.** **2008a**. Macrophytography of cultivated liriopogons and genera delineation. HortTechnology. 18: 334–342.

**Fantz, P. R.** **2008b**. Species of *Liriope* cultivated in the southeastern United States. HortTechnology. 18: 343–348.

**Fantz, P. R.** **2009**. Names and species of *Ophiopogon* cultivated in the southeastern United States. HortTechnology. 19: 385–394.

**Fantz, P. R., D. Carey, T. Avent, and J. Lattier**. **2015**. Inventory, descriptions, and keys to segregation and identification of liriopogons cultivated in the southeastern United States. HortScience. 50: 957–993.

**Freitas-Astúa, J., L. Moreira, C. Rivera, C. M. Rodrı́guez, and E. W. Kitajima**. **2002**. First report of orchid fleck virus in Costa Rica. Plant Disease. 86: 1402–1402.

**García-Escamilla, P., Y. Duran-Trujillo, G. Otero-Colina, G. Valdovinos-Ponce, Ma. T. Santillán-Galicia, C. F. Ortiz-Garcı́a, J. J. Velázquez-Monreal, and S. Sánchez-Soto**. **2018**. Transmission of viruses associated with cytoplasmic and nuclear leprosis symptoms by *Brevipalpus yothersi* and *B. californicus*. Tropical Plant Pathology. 43: 69–77.

**Gibbs, A.** **2000**. Viruses of orchids in Australia; their identification, biology and control. The Australian Orchid Rev. 65: 10–21.

**Kitajima, E. W., A. Blumenschein, and A. S. Costa**. **1974**. Rodlike particles associated with ringspot symptoms in several orchid species in Brazil. Journal of Phytopathology. 81: 280–286.

**Kitajima, E. W., C. M. Chagas, R. Harakava, R. F. Calegario, J. Freitas-Astúa, J. C. V. Rodrigues, and C. C. Childers**. **2011**. Citrus leprosis in Florida, USA, appears to have been caused by the nuclear type of citrus leprosis virus (CilLV-N). Virus Reviews & Research. 16.

**Kitajima, E. W., H. Kondo, A. Mackenzie, J. A. M. Rezende, R. Gioria, A. Gibbs, and T. Tamada**. **2001**. Comparative cytopathology and immunocytochemistry of Japanese, Australian and Brazilian isolates of orchid fleck virus. Journal of General Plant Pathology. 67: 231–237.

**Kitajima, E. W., J. C. V. Rodrigues, and J. Freitas-Astua**. **2010**. An annotated list of ornamentals naturally found infected by *Brevipalpus* mite-transmitted viruses. Scientia Agricola. 67: 348–371.

**Knorr, L. C.** **1968**. Studies on the etiology of leprosis in citrus. *In* International Organization of Citrus Virologists Conference Proceedings.

**Ko, N.-J., F. W. Zettler, J. R. Edwardson, and R. G. Christie**. **1985**. Light microscopic techniques for detecting orchid viruses. Acta Horticulturae. 241–254.

**Kondo, H., T. Maeda, Y. Shirako, and T. Tamada**. **2006**. Orchid fleck virus is a rhabdovirus with an unusual bipartite genome. Journal of General Virology. 87: 2413–2421.

**Kondo, H., T. Maeda, and T. Tamada**. **2003**. Orchid fleck virus: *Brevipalpus californicus* mite transmission, biological properties and genome structure. Experimental and Applied Acarology. 30: 215–223.

**Kondo, H., K. Maruyama, S. Chiba, I. B. Andika, and N. Suzuki**. **2014**. Transcriptional mapping of the messenger and leader RNAs of orchid fleck virus, a bisegmented negative-strand RNA virus. Virology. 452-453: 166–174.

**Kubo, K. S., J. Freitas-Astúa, M. A. Machado, and E. W. Kitajima**. **2009**. Orchid fleck symptoms may be caused naturally by two different viruses transmitted by *Brevipalpus*. Journal of General Plant Pathology. 75: 250–255.

**Lattier, J. D., T. G. Ranney, P. R. Fantz, and T. Avent**. **2014**. Identification, nomenclature, genome sizes, and ploidy levels of *Liriope* and *Ophiopogon* taxa. HortScience. 49: 145–151.

**Leeuwen, T. V., L. Tirry, A. Yamamoto, R. Nauen, and W. Dermauw**. **2015**. The economic importance of acaricides in the control of phytophagous mites and an update on recent acaricide mode of action research. Pesticide Biochemistry and Physiology. 121: 12–21.

**León, G. P.-P. de, and S. A. Nadler**. **2010**. What we don’t recognize can hurt us: A plea for awareness about cryptic species. Journal of Parasitology. 96: 453–464.

**Lesemann, D., and J. Begtrup**. **1971**. Elektronenmikroskopischer nachweis eines bazilliformen virus in *Phalaenopsis*. Journal of Phytopathology. 71: 257–269.

**Lesemann, D., and S. Doraiswamy**. **1975**. Bullet-shaped virus-like particles in chlorotic and necrotic leaf lesions of orchids. Journal of Phytopathology. 83: 27–39.

**Maeda, T.** **1998**. Evidence that orchid fleck virus is efficiently transmitted in a persistent manner by the mite *Brevipalpus californicus*. Abstr., 7th Int. Cong. Plant Pathol. 3.

**Magalhães, B. P., J. C. V. Rodrigues, D. G. Boucias, and C. C. Childers**. **2005**. Pathogenicity of *metarhizium anisopliae* var. Acridum to the false spider mite *brevipalpus phoenicis* (acari: tenuipalpidae). Florida Entomologist. 88: 195–198.

**Masiero, E., D. Banik, J. Abson, P. Greene, A. Slater, and T. Sgamma**. **2020**. Molecular verification of the UK national collection of cultivated *Liriope* and *Ophiopogon* plants. Plants. 9: 558.

**Mcharo, M., E. Bush, D. L. Bonte, C. Broussard, and L. Urbatsch**. **2003**. Molecular and morphological investigation of ornamental liriopogons. Journal of the American Society for Horticultural Science. 128: 575–577.

**Mei, Y., N. Bejerman, K. S. Crew, N. McCaffrey, and R. G. Dietzgen**. **2016**. First report of orchid fleck virus in lilyturf (*Liriope spicata*) in Australia. Plant Disease. 100: 1028–1028.

**Messing, R., and J. Brodeur**. **2017**. Current challenges to the implementation of classical biological control. BioControl. 63: 1–9.

**Nesom, G. L.** **2010**. Overview of *liriope* and *ophiopogon* (ruscaceae) naturalized and commonly cultivated in the USA. Phytoneuron. 56: 1–31.

**Omoto, C., E. B. Alves, and P. C. Ribeiro**. **2000**. Detecção e monitoramento da resistência de *brevipalpus phoenicis* (Geijskes) (acari: Tenuipalpidae) do dicofol. Anais da Sociedade Entomológica do Brasil. 29: 757–764.

**Pearson, M. N., G. V. H. Jackson, S. P. Pone, and R. L. J. Howitt**. **1993**. Vanilla viruses in the South Pacific. Plant Pathology. 42: 127–131.

**Peng, D. W., G. H. Zheng, Q. X. Tong, Z. Z. Zheng, and Y. L. Ming**. **2017**. First report of orchid fleck dichorhavirus from *cymbidium* sp. In China. Plant Disease. 101: 514–514.

**Peng, D. W., G. H. Zheng, Z. Z. Zheng, Q. X. Tong, and Y. L. Ming**. **2013**. Orchid fleck virus: An unclassified bipartite, negative-sense RNA plant virus. Archives of Virology. 158: 313–323.

**Petzold, H.** **1971**. Der elektronenmikroskopische nachweis eines bacilliformen virus an blattfleckenkranken *Dendrobien*. Journal of Phytopathology. 70: 43–52.

**Ramos-González, P. L., H. Sarubbi-Orue, L. Gonzales-Segnana, C. Chabi-Jesus, J. Freitas-Astúa, and E. W. Kitajima**. **2015**. Orchid fleck virus infecting orchids in Paraguay: First report and use of degenerate primers for its detection. Journal of Phytopathology. 164: 342–347.

**Rodrigues, J. C. V., and M. A. Machado**. **2000**. Virus-*Brevipalpus*-plant relationships on citrus leprosis pathosystems. Proc. Int. Soc. Citriculture Congr. 3–7.

**Roy, A., J. S. Hartung, W. L. Schneider, J. Shao, G. Leon, M. J. Melzer, J. J. Beard, G. Otero-Colina, G. R. Bauchan, R. Ochoa, and R. H. Brlansky**. **2015**. Role bending: Complex relationships between viruses, hosts, and vectors related to citrus leprosis, an emerging disease. Phytopathology. 105: 1013–1025.

**Roy, A., A. L. Stone, G. Otero-Colina, G. Wei, R. H. Brlansky, R. Ochoa, G. Bauchan, W. L. Schneider, M. K. Nakhla, and J. S. Hartung**. **2020**. Reassortment of genome segments creates stable lineages among strains of orchid fleck virus infecting citrus in Mexico. Phytopathology. 110: 106–120.

**Roy, A., A. Stone, G. Otero-Colina, G. Wei, N. Choudhary, D. Achor, J. Shao, L. Levy, M. K. Nakhla, C. R. Hollingsworth, J. S. Hartung, W. L. Schneider, and R. H. Brlansky**. **2013**. Genome assembly of citrus leprosis virus nuclear type reveals a close association with orchid fleck virus. Genome Announcements. 1.

**Sauvêtre, P., E. Veniant, G. Croq, A. D. Tassi, E. W. Kitajima, C. Chabi-Jesus, P. L. Ramos-González, J. Freitas-Astúa, and D. Navia**. **2018**. First report of orchid fleck virus in the orchid collection of jardin du luxembourg, Paris, France. Plant Disease. 102: 2670–2670.

**Skoracka, A., S. Magalhães, B. G. Rector, and L. Kuczyński**. **2015**. Cryptic speciation in the acari: A function of species lifestyles or our ability to separate species? Experimental and Applied Acarology. 67: 165–182.

**Suckling, D. M., L. D. Stringer, A. E. A. Stephens, B. Woods, D. G. Williams, G. Baker, and A. M. El-Sayed**. **2013**. From integrated pest management to integrated pest eradication: Technologies and future needs. Pest Management Science. 70: 179–189.

**Vechia, J. F. D., M. C. Ferreira, and D. J. Andrade**. **2018**. Interaction of spirodiclofen with insecticides for the control of *Brevipalpus yothersii* in citrus. Pest Management Science. 74: 2438–2443.

**Walker, P. J., K. R. Blasdell, C. H. Calisher, R. G. Dietzgen, H. Kondo, G. Kurath, B. Longdon, D. M. Stone, R. B. Tesh, N. Tordo, N. Vasilakis, and A. E. Whitfield**. **2018**. ICTV virus taxonomy profile: rhabdoviridae. Journal of General Virology. 99: 447–448.

**Wang, G.-Y., Y. Meng, J.-L. Huang, and Y.-P. Yang**. **2014**. Molecular phylogeny of *Ophiopogon* (asparagaceae) inferred from nuclear and plastid DNA sequences. Systematic Botany. 39: 776–784.

**Zheng, G. H., Z. Z. Zheng, Q. X. Tong, Y. L. Ming, and others**. **2013**. Orchid fleck virus: An unclassified bipartite, negative-sense rna plant virus. Archives of virology. 158: 313–323.

**Zhou, Q., J. Zhou, J. Chen, and X. Wang**. **2009**. Karyotype analysis of medicinal plant *Liriope spicata* var. *prolifera* (liliaceae). Biologia. 64.

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### Figure captions

Fig. 1: Variety of symptoms expressed by plants infected with Orchid fleck dichorhavirus: (a) ringspot symptoms on *Liriope gigantea* (b) chlorotic flecking on *Aspidistra elatior*.

### Figures

